

Genome editing made easy: Top 10 tips & tricks



TAL effector-mediated genome editing

- 1** Design and test at least 2–3 pairs of transcription activator-like (TAL) effectors per gene

 - Testing multiple constructs increases the chance of obtaining a highly specific and active TAL effector pair.
- 2** Design the TAL pair to cleave the DNA as close as possible to the desired position

 - Cleavage efficiency may vary based on the accessibility of the target loci.
 - Target accessibility could be affected by chromatin structure and DNA methylation.
 - [GeneArt™ PerfectMatch TALs](#) can be designed to target any desired locus, since there is no 5′ base restriction (5′ base at target site can be A, T, G, or C).
- 3** Design TAL repeats to target 18 or 24 bp of DNA sequence, and design TAL pair targets with 16 bp of spacing in between; then add one N (A, T, G, or C) to the 5′ end of each target sequence

 - GeneArt PerfectMatch TALs are available with 18 or 24 repeats to target 18- or 24-bp DNA-binding sequences, respectively. To order PerfectMatch TALs, a 5′ N has to be included in the target sequences. N could be A, T, G, or C.
 - The optimized spacing between forward TAL target and reverse TAL target is 16 bp.
- 4** GC content should be distributed throughout the target site when possible

 - Avoid long runs of As and Ts, especially at the 5′ end of the binding site.

- 5** Select the TAL vector of interest
- Choose the N-TAL Fok1 CMV expression vector to drive high-level expression of the TAL in mammalian systems. It can be used directly without extra subcloning.
 - Choose the N-TAL Fok1 entry vector (Gateway™-adapted vector) for easy transfer of your target-specific TAL to a destination vector designed for expression in your specific cell type.

- 6** Validate TALs with the GeneArt™ Genomic Cleavage Selection Kit and GeneArt™ Genomic Cleavage Detection Kit
- TAL function can be validated using the GeneArt Genomic Cleavage Selection Kit and the GeneArt Genomic Cleavage Detection Kit prior to performing full-scale genome engineering experiments.

- 7** Optimize transfection conditions
- High transfection efficiency is important for efficient genome modification with TALs.
 - Use our [transfection reagent selection guide](#) to find the best transfection reagent for your cell line and optimize transfection conditions according to cell type.
 - Optimize the amount of TAL pair for your cell type and culture condition. A good starting point is 750 ng of DNA or 500 ng of mRNA for each TAL pair, per well, in 12-well format.

- 8** Use mRNA instead of DNA
- To circumvent random integration and cell type-specific promoter requirements, use TAL mRNA instead of TAL expression plasmid.
 - Order ready-to-use TAL mRNA from our [Custom Services team](#) or make your own.
 - GeneArt PerfectMatch TAL vectors (pDEST40 or N-TAL CMV) have a T7 promoter upstream of the TAL Fok1 sequence, which can be used for *in vitro* transcription of TAL mRNA using the [mMESSAGE mMACHINE™ T7 ULTRA Transcription Kit](#). We recommend [Lipofectamine™ MessengerMAX™ reagent](#) for mRNA transfection.

- 9** Test modification efficiency with the GeneArt Genomic Cleavage Detection Kit
- Refer to the [GeneArt Genomic Cleavage Detection Kit](#) manual for analyzing genome modification efficiency after transfection.

- 10** Enrich modified cells with the GeneArt Genomic Cleavage Selection Kit
- In case of low transfection efficiency, modified cells can be enriched using the [GeneArt Genomic Cleavage Selection Kit](#).

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