

Protocol Outline

- Plate cells so they will be 70–90% confluent at the time of transfection.
- Prepare plasmid DNA-lipid complexes (recommend 2 doses of lipid).
- Add DNA-lipid complexes to cells.

Transfection Amounts

Component	96-well	24-well	6-well
DNA per well	100 ng	500 ng	2500 ng
Enhancer™ Reagent per well	0.2 µg	1 µg	5 µg
Lipofectamine® 3000 Reagent per well	0.15 and 0.3 µL	0.75 and 1.5 µL	3.75 and 7.5 µL

See the table inside of the foldout to view a typical culturing procedure along with a diagram.

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	Package Contents	<p>Catalog Numbers</p> <p>L3000001 L3000008 L3000015 L3000075 L3000150</p> <p>Size: 0.1 mL 0.75 mL 1.5 mL 5 × 1.5 mL 15 mL</p>
	Storage Conditions	<ul style="list-style-type: none"> Store at 4°C (do not freeze).
	Required Materials	<ul style="list-style-type: none"> Plasmid DNA (0.5–5 µg/µL stock) Opti-MEM® Reduced Serum Medium Microcentrifuge tubes
	Timing	<p>Preparation: 10 minutes Incubation: 5 minutes Final Incubation: 1–3 days</p>
	Selection Guide	<p>Lipofectamine® Reagents Go online to view related products.</p>
	Product Description	<ul style="list-style-type: none"> Lipofectamine® 3000 Reagent is a proprietary formulation for transfecting nucleic acids into a wide range of eukaryotic cells and especially designed for hard to transfect cells Make DNA-Lipofectamine® 3000 complexes in serum-free medium such as Opti-MEM® Reduced Serum Medium and add directly to cells in culture medium, in the presence or absence of serum/antibiotic. It is not necessary to remove complexes or change/add medium after transfection. The amount of Lipofectamine® 3000 Reagent required for successful transfection varies. Start any new transfection by testing the recommended two concentrations of Lipofectamine® 3000 Reagent to determine an optimum amount.
	Important Guidelines	
	Online Resources	<p>Visit our product page for additional information and protocols. For support, visit www.lifetechnologies.com/support.</p>

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Lipofectamine® 3000 Transfection Reagents Protocol

Transfect cells according to the following table. Use the indicated volume of DNA and P3000™ Reagent with each of the 2 volumes of Lipofectamine® 3000. **Each reaction mix is given per well and accounts for pipetting variations.**

Timeline		Steps	Procedure Details				
Day 0	1	 Seed cells to be 70–90% confluent at transfection	Component	96-well	24-well	6-well	
	2	 Vortex 2–3 sec	Adherent cells	1–4 × 10 ⁴	0.5–2 × 10 ⁵	0.25–1 × 10 ⁶	
			Dilute Lipofectamine® 3000 Reagent in Opti-MEM® Medium – Mix well	Opti-MEM® Medium	5 µL × 2	25 µL × 2	125 µL × 2
Day 1	3		Lipofectamine® 3000 Reagent	0.15 and 0.3 µL	0.75 and 1.5 µL	3.75 and 7.5 µL	
			Prepare master mix of DNA by diluting DNA in Opti-MEM® Medium, then add P3000™ Reagent – Mix well	Opti-MEM® Medium	10 µL	50 µL	250 µL
			DNA (0.5–5 µg/µL)	0.2 µg	1 µg	5 µg	
	4		P3000™ Reagent (2 µL/µg DNA)	0.4 µL	2 µL	10 µL	
			Add diluted DNA to each tube of diluted Lipofectamine® 3000 Reagent (1:1 ratio)	Diluted DNA (with P3000™ Reagent)	5 µL	25 µL	125 µL
	5		Incubate	Incubate for 5 minutes at room temperature.			
	6		Add DNA-lipid complex to cells	Component (per well)	96-well	24-well	6-well
DNA-lipid complex				10 µL	50 µL	250 µL	
DNA amount				100 ng	500 ng	2500 ng	
Day 2–4	7		Enhancer amount	0.2 µL	1 µL	5 µL	
			Lipofectamine®3000 Reagent used	0.15 and 0.3 µL	0.75 and 1.5 µL	3.75 and 7.5 µL	
			Visualize/analyze transfected cells	Incubate cells for 2–4 days at 37°C. Then, analyze transfected cells.			