

LRRK2 Inhibitor Evaluation and Profiling in Both Biochemical and Cellular HTS Formats by SelectScreen® Screening Services

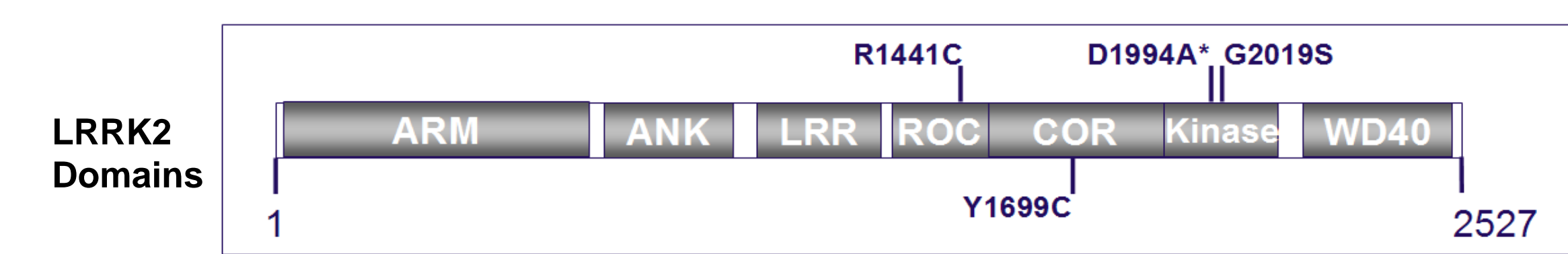


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ABSTRACT

Parkinson's Disease (PD) is a progressive neurological disorder that results from degeneration of neurons in a region of the brain that controls movement, impacting an estimated 6.3 million people worldwide. To date, mutations in the leucine-rich repeat kinase 2 (LRRK2) gene are the most common genetic cause of PD. LRRK2 is a protein kinase now being aggressively pursued as a target for PD therapeutics. However, studies of LRRK2 function and inhibition have been hampered by a lack of tools to express LRRK2 in diverse cell types as well as high-throughput biochemical and cell based assays. To address these needs, we have developed BacMam particles to express both untagged and green fluorescent protein (GFP) tagged full-length LRRK2 (WT and disease relevant mutants). BacMam technology has enabled higher levels of expression of LRRK2 in both workhorse cell lines (e.g. U-2 OS, HEK293T) and cells more physiologically relevant to neuroscience such as neuroblastoma SH-SY5Y and primary astrocytes compared to standard lipid transfection. Furthermore, the ability to express high levels of LRRK2 in these systems has enabled modular high-throughput assays for full-length LRRK2. Utilizing BacMam expression and LanthaScreen® technology, biochemical assays to measure inhibitor binding and cellular assays to measure inhibitor effects on kinase activity were developed and implemented by SelectScreen® Screening Services. Together these technologies will advance the field of PD research and therapeutics by addressing the currently unmet need for modular over expression, inhibitor binding assays and cellular kinase assays for full-length LRRK2.

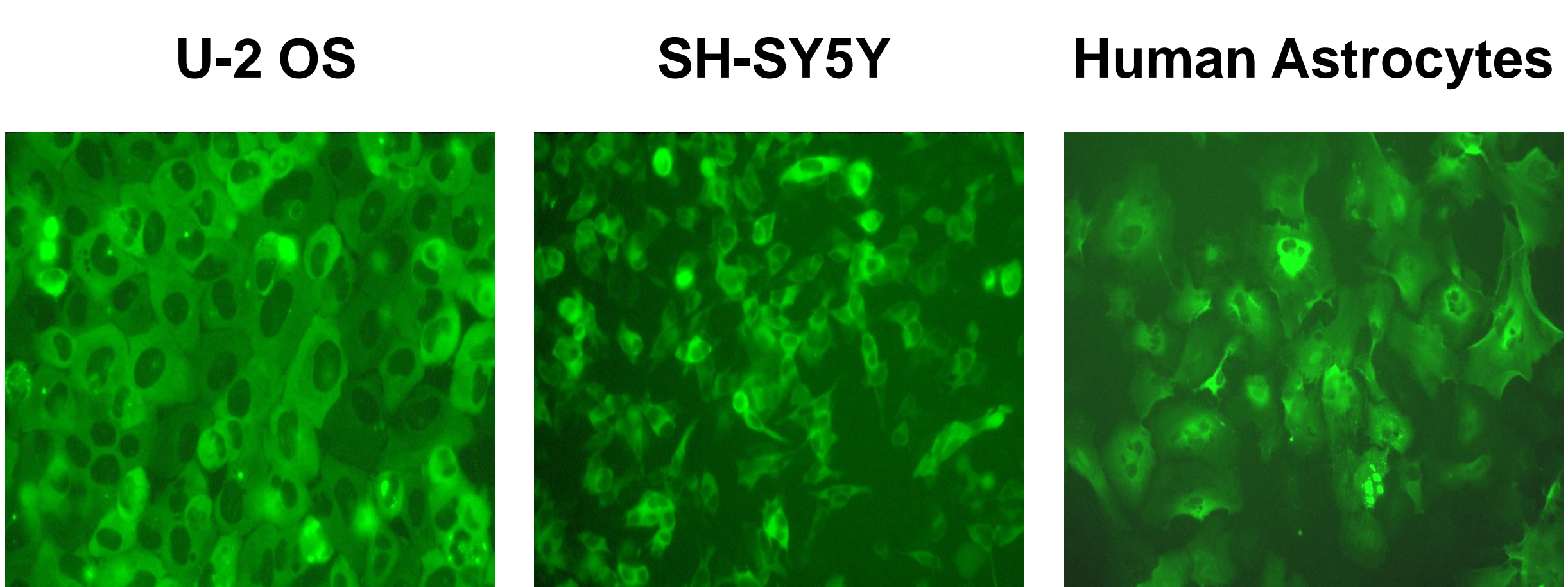
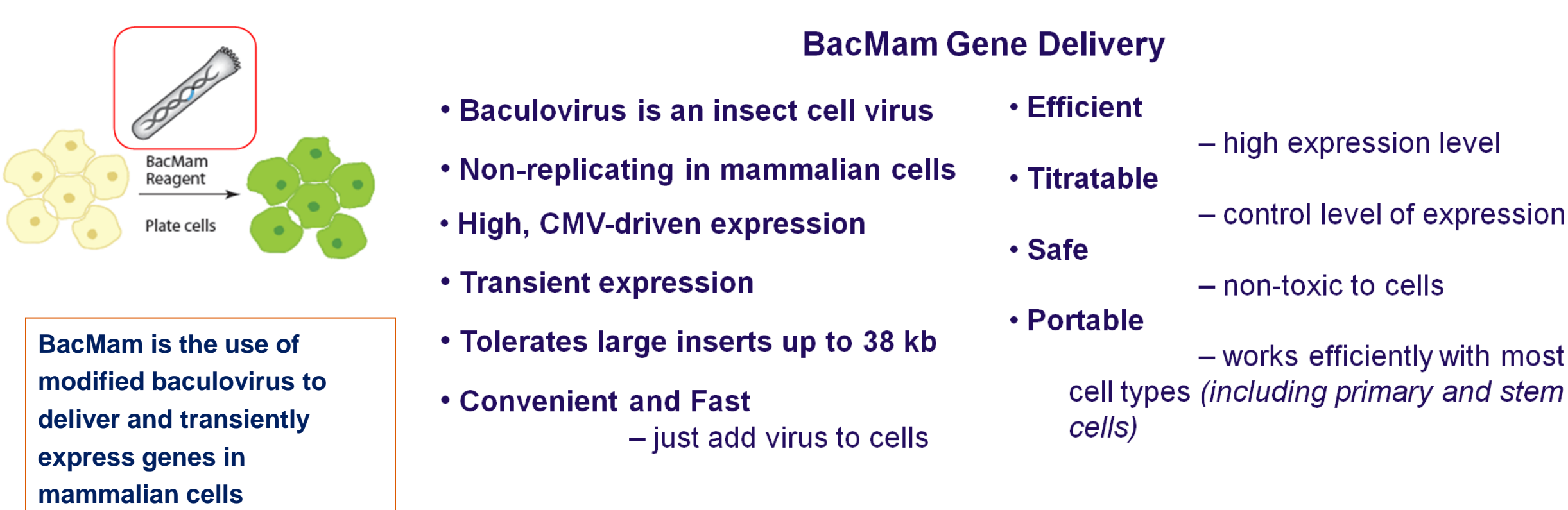
Figure 1. The Role of LRRK2 in Parkinson's Disease



The normal function of LRRK2 is unknown but it is expressed in many tissues and organs including the brain. LRRK2 is reported to be associated with microtubules, cytoskeleton rearrangements, synaptic vesicle trafficking, neurite outgrowth and translational control. LRRK2 is a large protein (2527 amino acid) belonging to the ROC family of proteins defined by the GTP-binding Ras of complex protein (ROC) and carboxy-terminal of Roc (COR) domains that confer weak GTPase activity. A serine/threonine protein kinase domain is positioned next to the ROC and COR domains completing the catalytic core of LRRK2. The catalytic core is flanked by leucine-rich repeats (LRR) on one side and a WD40 domain on the opposite side, both domains are thought to function as protein-protein interaction domains. Over 40 mutations of LRRK2 have been reported but only mutations in the catalytic core of LRRK2 segregate with PD in a Mendelian fashion suggesting the enzymatic functions of LRRK2 play a key role in disease pathogenesis. Mutations in the ROC GTPase domain (R1441C, R1441G, R1441H) and the COR domain (Y1699C) are reported to reduce GTPase activity in in-vitro studies. The most frequent PD associated LRRK2 mutation G2019S is located in the activation loop of the kinase domain resulting in increased kinase activity and reported to cause neurotoxicity in cultured cells.

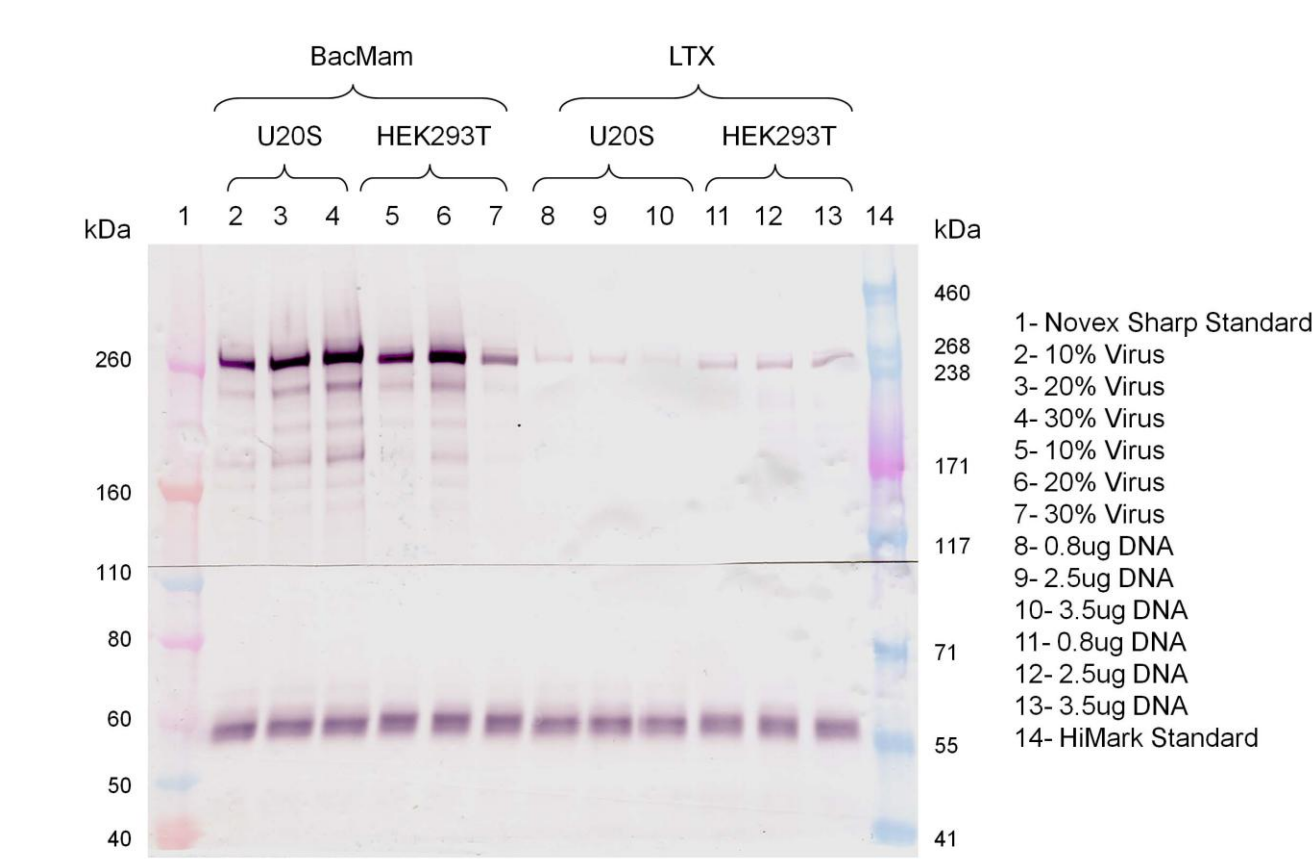
RESULTS

Figure 2. BacMam Enabled Expression of Parkinson's Disease Drug Target LRRK2



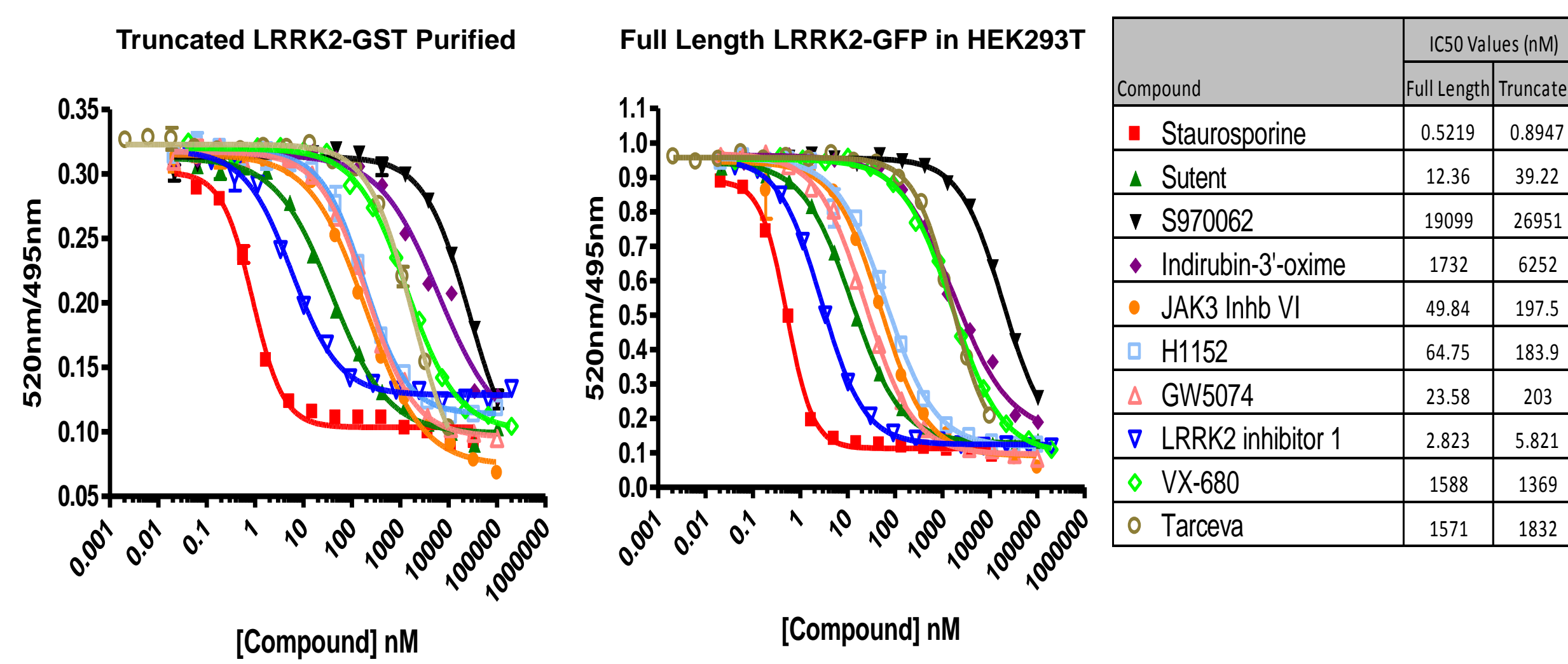
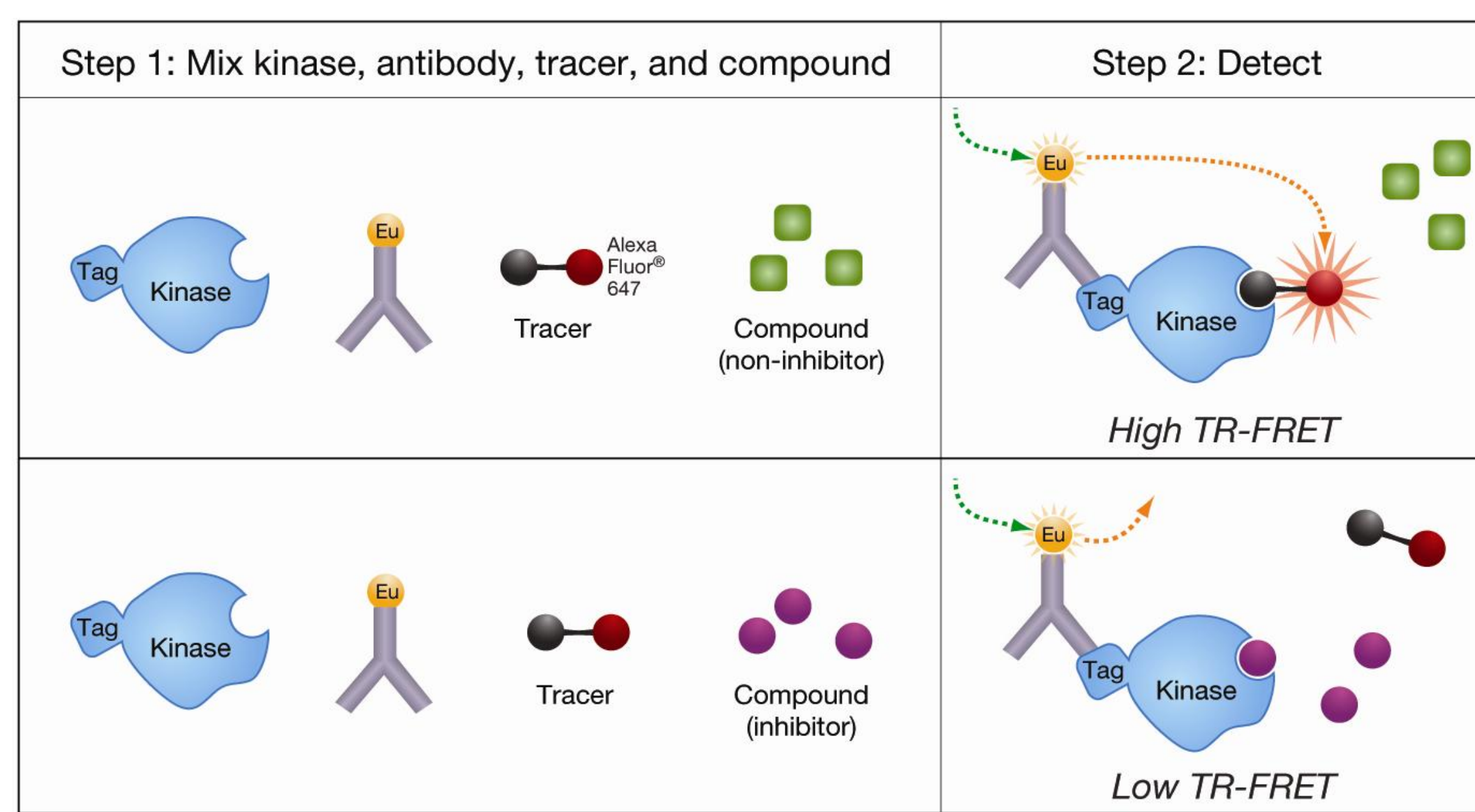
U-2 OS cells, SH-SY5Y Cells and human astrocytes (N7805100) were transduced with BacMam viruses of GFP tagged LRRK2 (A14170) and mutants. Cells were exposed to 20% virus for 24 hours, media was exchanged and cells were allowed to incubate an additional 24 hours before imaging by fluorescent microscopy.

Figure 3. Expression Level Comparison of BacMam VS Lipid Transfection



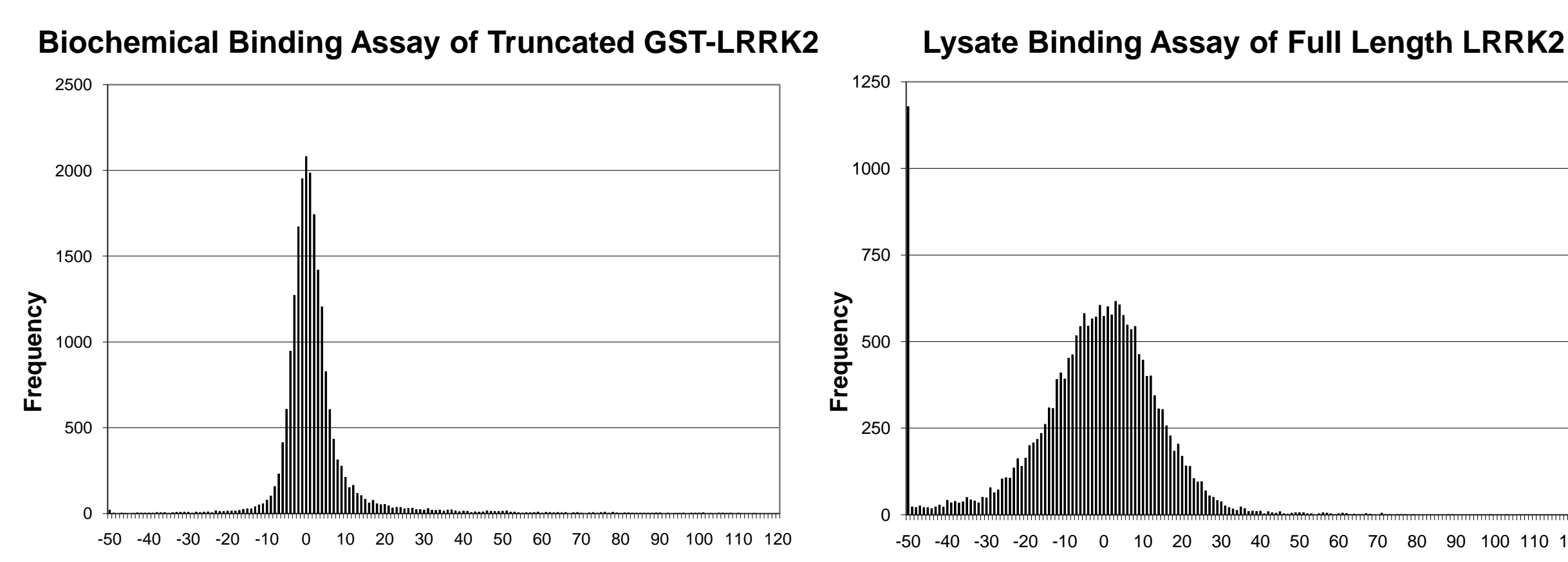
Expression level comparison of BacMam versus Lipofectamine LTX (with plus reagent) was tested in U-2 OS and HEK293T cells under optimized levels of full length LRRK2 were assessed by western blot (rabbit anti-LRRK2 - Cell Signaling Technologies). Sample loading was normalized by Bradford assays and confirmed by anti-AKT (Cell Signaling Technologies) blot (lower bands).

Figure 4. Kinase Binding Assay Utilizing Lysates from BacMam LRRK2 Transduced HEK293T Cells



BacMam LRRK2-GFP reagent was used to transduce HEK293T and generate a cellular lysate containing full length LRRK2. A lysate based Kinase Binding assay was developed using the full length LRRK2 lysate and compared to purified truncated LRRK2. Optimal lysate and tracer concentration were determined. 5nM truncated kinase (PV4873) or 3nM full length kinase (A14171), 2 nM Eu-anti GFP antibody (A14173) and 20 nM Tracer 236 (PV5592) were used to assay binding of several kinase inhibitors to LRRK2.

Figure 5. Enamine Library Screen Using Full Length LRRK2 Kinase Binding Assay



Percent Inhibition	Truncated LRRK2 (970-2527)	Full Length LRRK2
>70% inhibition	93/20,155 (0.46%)	12/20,155 (0.06%)
>30% inhibition	475/20,155 (2.36%)	270/20,155 (1.34%)
Average Z'-factor	0.80	0.55

Top Library Hits For LRRK2 Followed-Up Using Multiple Assay Formats

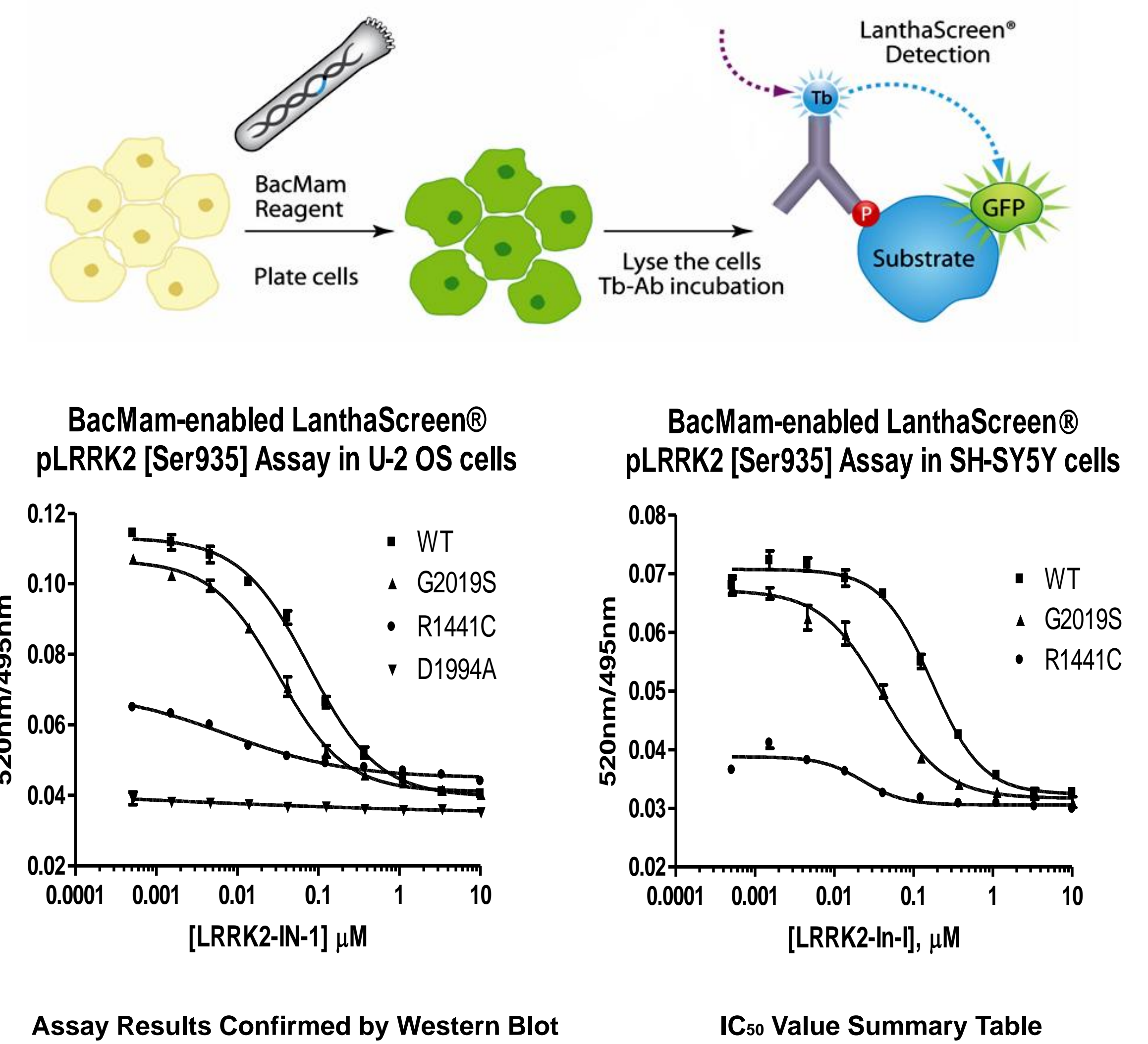
IC50 (µM)	Cellular			Biochemical Binding								Biochemical Activity	
	SH-SY5Y cells	U-2 OS cells	FL Lysate-based binding assay	Truncated, purified biochemical assay									
Compound Name	G2019S	WT	G2019S	WT	R1441C	G2019S	WT	R1441C	D1994A	G2019S	WT	G2019S	WT
LRRK2-IN-1	0.094	0.22	0.055	0.11	0.005	0.005	0.005	0.003	0.002	0.003	0.002	0.004	0.02
JAK3 Inhb. VI	0.73	1.2	0.21	1.4	0.23	0.2	nd	0.04	0.05	nd	nd	nd	nd
Indinubin-3-oxime	3.8*	>10	3.9*	1.9*	9.3	6.2	nd	2.1	1.7	nd	nd	nd	nd
Compd 2	>20	>20	>20	>20	0.13	0.15	1.9	3*	5.4*	>20	>20	>20	>20
Compd 5	>20	>20	>20	>20	3.3	1.8	7.2	1.2	1	1.2	1.1	1.3	>20
Compd 6	0.53	0.76	0.2	0.31	0.09	0.06	0.06	0.025	0.012	0.017	0.011	0.029	0.49
Compd 10	>10	>10	0.37*	0.55*	2	1.9	4.2	0.09	0.11	0.21	0.11	0.23	1.3*
Compd 17	>20	>20	20.9*	>20	0.84	0.8	0.59	0.39	0.26	0.34	0.24	0.52	1.8
Compd 25	0.17	0.88	0.99	1.22	0.03	0.19	0.12	0.01	0.019	0.028	0.014	0.053	0.11
Compd 27	nd	nd	0.44	1.29	0.25	0.274	0.6	0.054	0.054	0.1	0.056	0.19	0.54
Compd 30	>20	>20	>20	>20	8.5	1.6	1.3	10.6*	18.3*	>20	>20	11.1*	>20

Top Library Hits Profiled Against Kinase Panel To Determine Compound Specificity

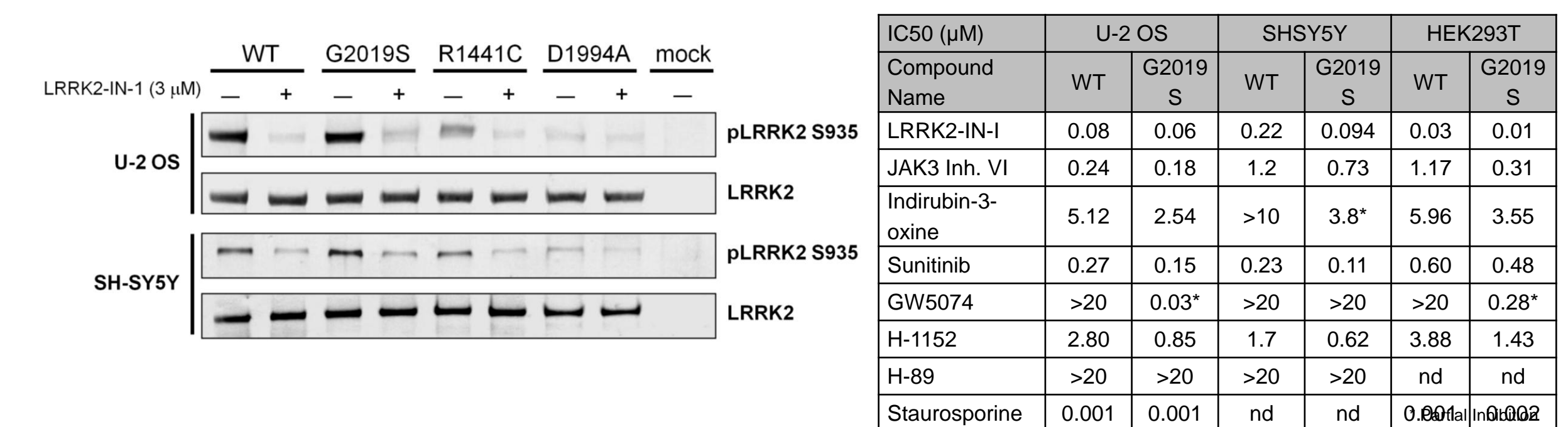
	LRRK2 IN-1	GW5074	Orpd 2	Orpd 5	Orpd 6	Orpd 10	Orpd 17	Orpd 15	Orpd 27	Orpd 30
>80% inhibition	7	126	0	216	52	8	115	14	221	5
20-80% inhibition	58	93	8	5	170	142	103	154	1	66

The lysate binding assay performed well in a high throughput screening (HTS) format when compared against the biochemical binding assay in a screen of the Enamine library - a drug like set of 20,155 compounds. Top hits from both screens were followed-up with IC50 curves in multiple assay formats including lysate and biochemical binding assay, cellular assay and biochemical activity assay. Selected compounds were also profiled by SelectScreen® Profiling Services against a panel of 223 kinases to assess compound specificity.

Figure 6. Cellular Assay for Monitoring the Phosphorylation of LRRK2 Ser935

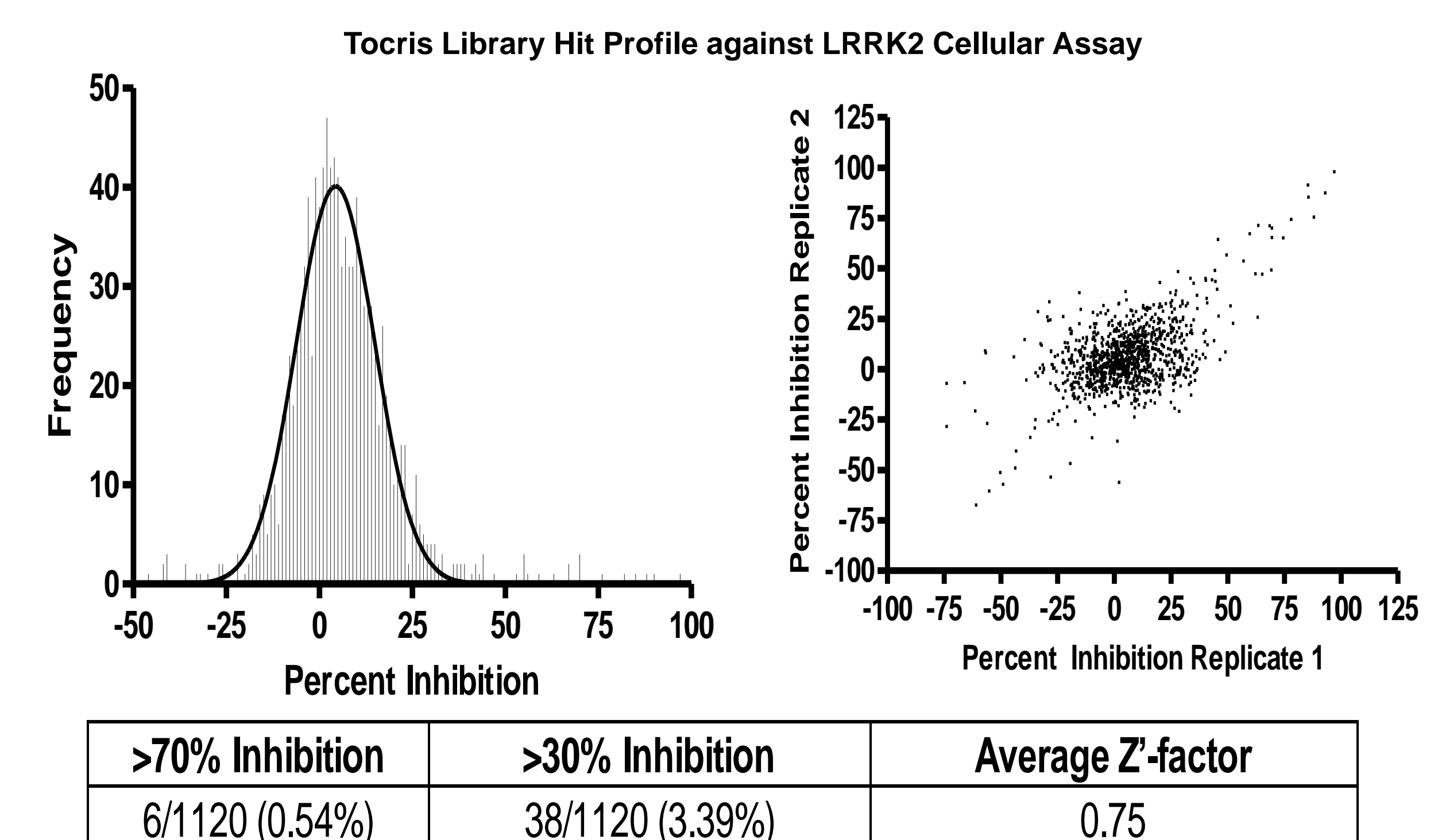


Assay Results Confirmed by Western Blot



Using Time-Resolved Förster Resonance Energy Transfer (TR-FRET) technology, we developed a high-throughput compatible homogenous cellular assay for monitoring the LRRK2 phosphorylation at Ser935. LRRK2-GFP fusion proteins were transiently expressed in a variety of cell backgrounds via BacMam gene delivery system. Cells were plated in 384-well assay plates. The phosphorylation at Ser935 in these cells was detected using a terbium labeled anti-Ser935 phosphorylation specific antibody that generates TR-FRET signals between terbium and GFP. Consistent with previous reports and our western blot results, wild type and G2019S LRRK2 are constitutively phosphorylated at Ser935 in cells measured by TR-FRET. The phosphorylation level was reduced for R1441C mutant and little could be detected for the kinase activity-dead mutant D1994A. The TR-FRET cellular assay was further validated with reported LRRK2 inhibitors including LRRK2-IN-1 and further confirmed that inhibition of LRRK2 kinase activity can reduce the phosphorylation level at Ser935.

Figure 7. Trocrist Mini Library Screen Using LRRK2 pSer935 Cellular Assay



To demonstrate the utility of the LRRK2 cellular assay in HTS applications a compound screen was performed using the Trocrist mini library - a collection of 1120 biologically active compounds.

CONCLUSIONS

An extensive set of tools have been developed to enable the discovery of kinase inhibitors for PD target LRRK2. These tools allow for HTS compatible screens in multiple formats that provide detailed mechanistic data on how compounds are binding LRRK2 and inhibiting kinase activity of LRRK2 and its mutants in biochemical and cellular environments. Additionally, all assays described are available through SelectScreen® Screening Services.

Useful Weblinks

- www.lifetechnologies.com/bindingassay
- www.lifetechnologies.com/lanthascreencellular
- www.lifetechnologies.com/kinaseprofiling