

# Phospho-S6 (Ser235, Ser236) Monoclonal Antibody (cupk43k), PE, eBioscience™

Product Details	
Size	25 Tests
Species Reactivity	Human, Mouse
Published Species	Mouse
Host/Isotope	Mouse / IgG1, kappa
Recommended Isotype Control	Mouse IgG1 kappa Isotype Control (P3.6.2.8.1), PE, eBioscience™
Class	Monoclonal
Type	Antibody
Clone	cupk43k
Conjugate	PE
Form	Liquid
Concentration	5 µL/Test
Purification	Affinity chromatography
Storage buffer	PBS, pH 7.2, with 0.1% gelatin, 0.2% BSA
Contains	0.09% sodium azide
Storage Conditions	4° C, store in dark, DO NOT FREEZE!
RRID	AB_2572666

Applications	Tested	Dilution	Published
Flow Cytometry (Flow)	✓	5 µL (0.125 µg)/test	1 Publication

## Product Specific Information

Description: This cupk43k monoclonal antibody recognizes human and mouse ribosomal protein S6 (also known as 40S ribosomal protein S6, phosphoprotein NP33, RPS6, RS6, S6) when phosphorylated on serine 235 (S235, human) and serine 236 (S236, mouse). Ribosomal protein S6 is a component of the 40S subunit of the ribosome and is phosphorylated at multiple sites following stimulation of cells by growth factors, tumor promoting agents, or mitogens. Phosphorylation of ribosomal protein S6 by p70S6K and PKDCD results in upregulation of the translation of RNA coding for proteins involved in cell cycle entry. Ribosomal protein S6 is dephosphorylated upon growth arrest.

The specificity of the cupk43k monoclonal antibody was determined by western blotting.

Applications Reported: This cupk43k antibody has been reported for use in intracellular staining followed by flow cytometric analysis.

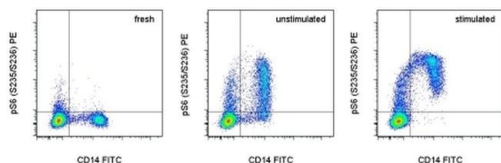
Applications Tested: This cupk43k antibody has been pre-titrated and tested by intracellular staining and flow cytometric analysis of stimulated normal human peripheral blood cells. This can be used at 5 µL (0.125 µg) per test. A test is defined as the amount (µg) of antibody that will stain a cell sample in a final volume of 100 µL. Cell number should be determined empirically but can range from 10<sup>5</sup> to 10<sup>8</sup> cells/test.

Use of Protocol A: Two-step protocol: intracellular (cytoplasmic) proteins allows for the greatest flexibility for detection of surface and intracellular (cytoplasmic) proteins. Use of Protocol B: One-step protocol: intracellular (nuclear) proteins is recommended for staining of transcription factors in conjunction with surface and phosphorylated intracellular (cytoplasmic) proteins. Protocol C: Two-step protocol: Fixation/Methanol allows for the greatest discrimination of phospho-specific signaling between unstimulated and stimulated samples, but with limitations on the ability to stain specific surface proteins (refer to "Clone Performance Following Fixation/Permeabilization" located in the Best Protocols Section under the Resources tab online). All Protocols can be found in the "Staining Intracellular Antigens for Flow Cytometry Protocol" located in the Best Protocols Section under the Resources tab online.

Excitation: 488-561 nm; Emission: 578 nm; Laser: Blue Laser, Green Laser, Yellow-Green Laser.

Filtration: 0.2 µm post-manufacturing filtered.

## Product Images For Phospho-S6 (Ser235, Ser236) Monoclonal Antibody (cupk43k), PE, eBioscience™



### Phospho-S6 (Ser235, Ser236) Antibody (12-9007-41) in Flow

Intracellular staining of freshly-harvested (left), unstimulated (middle), or 30-minute LPS-stimulated (right) normal human peripheral blood cells with Anti-Human CD14 FITC (Product # 11-0149-42) and Anti-Human phospho-S6 (S235/S236) PE, using the Intracellular Fixation and Permeabilization Buffer Set (Product # 88-8824-00) and protocol. Cells in the lymphocyte/monocyte gate were used for analysis.

## 1 Reference

### Flow Cytometry (1)

The Journal of experimental medicine

#### PTEN drives Th17 cell differentiation by preventing IL-2 production.

"12-9007 was used in Flow cytometry/Cell sorting to show that PTEN plays a key role in Th17 cell differentiation by blocking IL-2 expression."

Authors: Kim HS, Jang SW, Lee W, Kim K, Sohn H, Hwang SS, Lee GR

**Species**  
Mouse

**Dilution**  
Not Cited

**Year**  
2017

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