FOXP3 Monoclonal Antibody (236A/E7), PE-Cyanine7, eBioscience™

Product Details	
Size	100 Tests
Species Reactivity	Human, Non-human primate, Rhesus monkey
Published Species	Human
Host/Isotype	Mouse / IgG1, kappa
Recommended Isotype Control	Mouse IgG1 kappa Isotype Control (P3.6.2.8.1), PE-Cyanine7, eBioscience™
Class	Monoclonal
Type	Antibody
Clone	236A/E7
Conjugate	PE-Cyanine7
Excitation/Emission Max	569/780 nm
Form	Liquid
Concentration	5 µL/Test
Purification	Affinity chromatography
Storage buffer	PBS, pH 7.2, with 0.2% BSA
Contains	0.09% sodium azide
Storage conditions	4° C, store in dark, DO NOT FREEZE!
RRID	AB_2573450

Applications	Tested Dilution	Publications
Immunohistochemistry (Paraffin) (IHC (P))	-	2 Publications
Flow Cytometry (Flow)	5 µL (0.125 µg)/test	12 Publications

Product Specific Information

Description: The 236A/E7 antibody reacts with human Foxp3 protein also known as FORKHEAD BOX P3, SCURFIN, and JM2. Foxp3, a 49-55 kDa protein, is a member of the forkhead/winged-helix family of transcriptional regulators, and was identified as the gene defective in 'scurfy' (sf) mice. Constitutive high expression of Foxp3 mRNA has been shown in CD4+CD25+ regulatory T cells (Treg cells), and ectopic expression of Foxp3 in CD4+CD25- cells imparts a Treg phenotype in these cells.

Intracellular staining and flow cytometric analysis of freshly isolated human peripheral blood mononuclear cells (PBMCs) with the 236A/E7 antibody using the Foxp3/Transcription Factor Staining Buffer Set (cat. 00-5523) and protocol reveals staining of the CD4+CD25bright population.

The epitope from 236A/E7 is different from that of PCH101 (cat. 72-5776). This antibody has also been shown to recognize rhesus macaque, sooty mangabey and cynomolgus macaque.

Applications Reported: This 236A/E7 antibody has been reported for use in intracellular staining followed by flow cytometric analysis.

Applications Tested: This 236A/E7 antibody has been pre-titrated and tested by intracellular staining and flow cytometric

analysis of normal human peripheral blood cells using the Foxp3/Transcription Factor Staining Buffer Set (cat. 00-5523) and protocol. Please refer to Best Protocols: Protocol B: One step protocol for (nuclear) intracellular proteins. This can be used at 5 µL (0.125 µg) per test. A test is defined as the amount (µg) of antibody that will stain a cell sample in a final volume of 100 µL. Cell number should be determined empirically but can range from 10^5 to 10^8 cells/test.

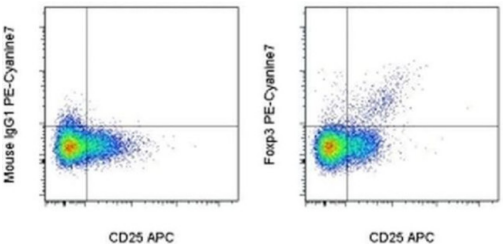
Light sensitivity: This tandem dye is sensitive photo-induced oxidation. Please protect this vial and stained samples from light.

Fixation: Samples can be stored in IC Fixation Buffer (cat. 00-8222) (100 µL cell sample + 100 µL IC Fixation Buffer) or 1-step Fix/Lyse Solution (cat. 00-5333) for up to 3 days in the dark at 4°C with minimal impact on brightness and FRET efficiency /compensation. Some generalizations regarding fluorophore performance after fixation can be made, but clone specific performance should be determined empirically.

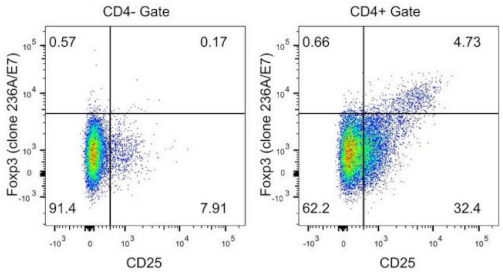
Excitation: 488-561 nm; Emission: 775 nm; Laser: Blue Laser, Green Laser, Yellow-Green Laser.

Filtration: 0.2 µm post-manufacturing filtered.

Product Images For FOXP3 Monoclonal Antibody (236A/E7), PE-Cyanine7, eBioscience™



FOXP3 Antibody (25-4777-42) in Flow
Intracellular staining of normal human peripheral blood cells with Anti-Human CD25 APC (Product # 17-0259-42) and Mouse IgG1 K Isotype Control PE-Cyanine7 (Product # 25-4714-80) (left) or Anti-Human Foxp3 PE-Cyanine7 (right) using the Foxp3/Transcription Factor Staining Buffer Set (Product # 00-5523-00) and protocol. Cells in the lymphocyte gate were used for analysis.



FOXP3 Antibody (25-4777-42)
Intracellular staining of human peripheral blood cells. As expected based on known expression patterns, Foxp3 clone 236A/E7 stains CD4+CD25+ T cells and does not stain CD4- cells or CD4+CD25- T cells. Details: Normal human peripheral blood cells were surface stained with CD4 (clone OKT4) and CD25 (clone BC96) followed by intracellular staining with Foxp3 (clone 236A/E7) using the Foxp3/Transcription Factor Staining Buffer Set and protocol. Lymphocytes in the CD4- (left) and CD4+ (right) gates were used for analysis. {RE}

14 References

Immunohistochemistry (Paraffin) (2)

The Journal of infectious diseases

CD4(+) regulatory T cells in a cynomolgus macaque model of Mycobacterium tuberculosis infection.

Authors: Green AM,Mattila JT,Bigbee CL,Bongers KS,Lin PL,Flynn JL

Year
2010

Journal of immunology (Baltimore, Md. : 1950)

Early resolution of acute immune activation and induction of PD-1 in SIV-infected sooty mangabeys distinguishes nonpathogenic from pathogenic infection in rhesus macaques.

Authors: Estes JD,Gordon SN,Zeng M,Chahroudi AM,Dunham RM,Staprans SI,Reilly CS,Silvestri G,Haase AT

Year
2008

Flow Cytometry (12)

Current issues in molecular biology

In Vitro Evidence of Differential Immunoregulatory Response between MDA-MB-231 and BT-474 Breast Cancer Cells Induced by Bone Marrow-Derived Mesenchymal Stromal Cells Conditioned Medium.

"25-4777-42 was used in Flow cytometry/Cell sorting to study the effect of the conditioned medium of human bone marrow-derived-MSCs (hBM-MSC-cm) on the immunoregulatory capability of MDA-MB-231 and BT-474 breast cancer cells."

Authors: Arenas-Luna VM,Montesinos JJ,Cortés-Morales VA,Navarro-Betancourt JR,Peralta-Ildefonso J,Cisneros B, Hernández-Gutiérrez S

Year
2022

Species
Human

European journal of immunology

Guidelines for the use of flow cytometry and cell sorting in immunological studies (third edition).

"25-4777-42 was used in Flow cytometry/Cell sorting to provide the key aspects to consider when performing flow cytometry experiments."

Authors: Cossarizza A,Chang HD,Radbruch A,Abrignani S,Addo R,Akdis M,Andrä I,Andreata F,Annunziato F,Arranz E, Bacher P,Bari S,Barnaba V,Barros-Martins J,Baumjohann D,Beccaria CG,Bernardo D,Boardman DA,Borger J,Böttcher C,Brockmann L,Burns M,Busch DH,Cameron G,Cammarata I,Cassotta A,Chang Y,Chirdo FG,Christakou E,Iin-Šain L, Cook L,Corbett AJ,Cornelis R,Cosmi L,Davey MS,De Biasi S,De Simone G,Del Zotto G,Delacher M,Di Rosa F,Di Santo J,Diefenbach A,Dong J,Dörner T,Dress RJ,Dutertre CA,Eckle SBG,Eede P,Evrard M,Falk CS,Feuerer M,Fillatreau S, Fiz-Lopez A,Follo M,Foulds GA,Fröbel J,Gagliani N,Galletti G,Gangaev A,Garbi N,Garrote JA,Geginat J,Gherardin NA, Gibellini L,Ginhoux F,Godfrey DI,Gruarin P,Haftmann C,Hansmann L,Harpur CM,Hayday AC,Heine G,Hernández DC, Herrmann M,Hoelsken O,Huang Q,Huber S,Huber JE,Huehn J,Hundemer M,Hwang WYK,Iannacone M,Ivison SM,Jäck HM,Jani PK,Keller B,Kessler N,Ketelaars S,Knop L,Knopf J,Koay HF,Kobow K,Kriegsmann K,Kristyanto H,Krueger A, Kuehne JF,Kunze-Schumacher H,Kvistborg P,Kwok I,Latorre D,Lenz D,Levings MK,Lino AC,Liotta F,Long HM,Lugli E, MacDonald KN,Maggi L,Maini MK,Mair F,Manta C,Manz RA,Mashreghi MF,Mazzoni A,McCluskey J,Mei HE,Melchers F, Melzer S,Mielenz D,Monin L,Moretta L,Multhoff G,Muñoz LE,Muñoz-Ruiz M,Muscate F,Natalini A,Neumann K,Ng LG, Niedobitek A,Niemz J,Almeida LN,Notarbartolo S,Ostendorf L,Pallett LJ,Patel AA,Percin GI,Peruzzi G,Pinti M,Pockley AG,Pracht K,Prinz I,Pujol-Autonell I,Pulvirenti N,Quatrini L,Quinn KM,Radbruch H,Rhys H,Rodrigo MB,Romagnani C, Saggau C,Sakaguchi S,Sallusto F,Sanderink L,Sandrock I,Schauer C,Scheffold A,Scherer HU,Schiemann M, Schildberg FA,Schober K,Schoen J,Schuh W,Schüler T,Schulz AR,Schulz S,Schulze J,Simonetti S,Singh J,Sitnik KM, Stark R,Starossom S,Stehle C,Szelinski F,Tan L,Tarnok A,Tornack J,Tree TIM,van Beek JJP,van de Veen W,van Gisbergen K,Vasco C,Verheyden NA,von Borstel A,Ward-Hartstonge KA,Warnatz K,Waskow C,Wiedemann A,Wilharm A,Wing J,Wirz O,Wittner J,Yang JHM,Yang J

Year
2021

Species
Human

View more Flow references on thermofisher.com

More applications with references on thermofisher.com