Donkey anti-Goat IgG (H+L) Cross-Adsorbed Secondary Antibody, Alexa Fluor™ 594

Product Details	
Size	1 mg
Species Reactivity	Goat
Host/Isotype	Donkey / IgG
Class	Polyclonal
Type	Secondary Antibody
Conjugate	Alexa Fluor™ 594
Excitation/Emission Max	590/618 nm
Immunogen	Gamma Immunoglobins Heavy and Light chains
Form	liquid
Concentration	2 mg/mL
Purification	purified
Storage buffer	PBS, pH 7.5
Contains	5mM sodium azide
Storage conditions	4° C, store in dark
RRID	AB_2534105

Applications	Tested Dilution	Publications
Immunohistochemistry (IHC)	1-10 µg/mL	0 Publication
Immunohistochemistry (Paraffin) (IHC (P))	-	0 Publication
Immunohistochemistry (PFA fixed) (IHC (PFA))	-	0 Publication
Immunohistochemistry (Frozen) (IHC (F))	-	0 Publication
Immunohistochemistry - Free Floating (IHC (Free))	-	0 Publication
Immunocytochemistry (ICC/IF)	1-10 µg/mL	0 Publication
Flow Cytometry (Flow)	1-10 µg/mL	-
Miscellaneous PubMed (Misc)	-	0 Publication

Product Specific Information

To minimize cross-reactivity, these donkey anti-goat IgG (H+L) whole secondary antibodies have been affinity purified and cross-adsorbed against rabbit, rat, mouse, and human IgG. Cross-adsorption or pre-adsorption is a purification step to increase specificity of the antibody resulting in higher sensitivity and less background staining. The secondary antibody solution is passed through a column matrix containing immobilized serum proteins from potentially cross-reactive species. Only the nonspecific-binding secondary antibodies are captured in the column, and the highly specific secondaries flow through. The benefits of this extra step are apparent in multiplexing/multicolor-staining experiments (e.g., flow cytometry) where there is potential cross-reactivity with other primary antibodies or in tissue/cell fluorescent staining experiments where there are may be the presence of endogenous immunoglobulins.

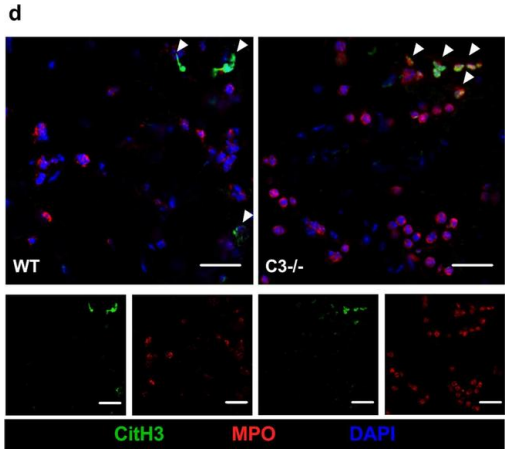
Alexa Fluor dyes are among the most trusted fluorescent dyes available today. Invitrogen™ Alexa Fluor 594 dye is a bright, red-fluorescent dye with excitation ideally suited to the 594 nm laser line. For stable signal generation in imaging and flow

cytometry, Alexa Fluor 594 dye is pH-insensitive over a wide molar range. Probes with high fluorescence quantum yield and high photostability allow detection of low-abundance biological structures with great sensitivity. Alexa Fluor 594 dye molecules can be attached to proteins at high molar ratios without significant self-quenching, enabling brighter conjugates and more sensitive detection. The degree of labeling for each conjugate is typically 2-8 fluorophore molecules per IgG molecule; the exact degree of labeling is indicated on the certificate of analysis for each product lot.

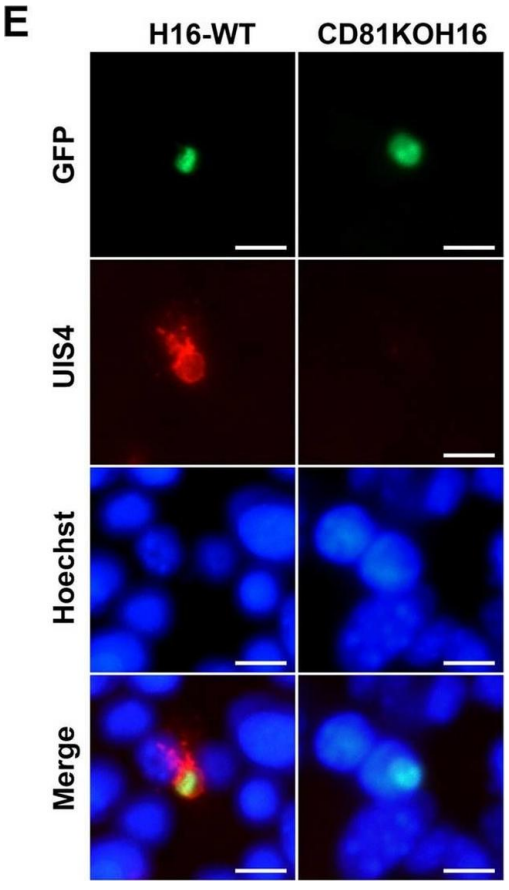
Using conjugate solutions: Centrifuge the protein conjugate solution briefly in a microcentrifuge before use; add only the supernatant to the experiment. This step will help eliminate any protein aggregates that may have formed during storage, thereby reducing nonspecific background staining. Because staining protocols vary with application, the appropriate dilution of antibody should be determined empirically. For the fluorophore-labeled antibodies a final concentration of 1-10 µg/mL should be satisfactory for most immunohistochemistry and flow cytometry applications.

Product will be shipped at Room Temperature.

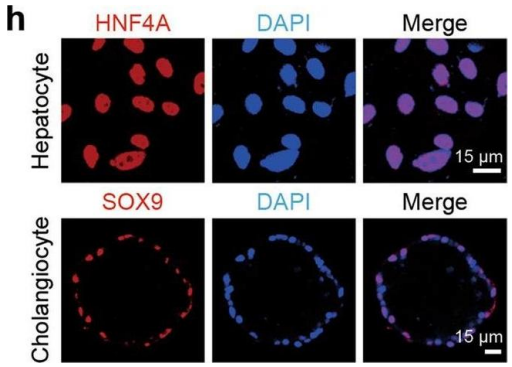
Product Images For Donkey anti-Goat IgG (H+L) Cross-Adsorbed Secondary Antibody, Alexa Fluor™ 594



Goat IgG (H+L) Cross-Adsorbed Secondary Antibody (A-11058) in ICC/IF
C3/ mice show a higher number of neutrophils and NETs. Scatter plots show (a) neutrophils (MPO+) per mm2, (b) neutrophil extracellular traps (MPO+/CitH3+) per mm2 and (c) the percentage of NETs/neutrophils in ischemic gastrocnemius muscles isolated 24 h after FAL. Data shown are means ± SEM, n = 5 per group, a defined ischemic area (0.86 mm2) of muscle tissue was analyzed per mouse. * p < 0.05, ns 0.05 (WT vs. C3/) by unpaired Student&quote;s t-test. (d) Representative immunofluorescence pictures of analyzed ischemic gastrocnemius muscles of WT (left) and C3/ mice (right). Cells were labeled with antibodies targeting MPO (red), CitH3 (green), and with DAPI (blue) to label nuclei. NETs (MPO+/CitH3+) are indicated by white arrowheads. Scale bars: 20 µm. Image collected and cropped by CiteAb from the following publication (https://pubmed.ncbi.nlm.nih.gov/34071589), licensed under a CC BY license.



Goat IgG (H+L) Cross-Adsorbed Secondary Antibody (A-11058) in ICC/IF
CRISPR-mediated inactivation of CD81 abrogates P. berghei infection in Hepa1-6 cells. (A) Hepa1-6 and CD81KOH16 cells were stained for surface CD81 with anti-CD81 MT81 monoclonal antibody and Alexa Fluor 488-conjugated secondary antibodies, before flow cytometry analysis. Histograms represent the fluorescence intensity of extracellular CD81 proteins for WT Hepa1-6 (blue) and CD81KOH16 cells (orange). The grey histogram represents cells stained with secondary antibodies only (Control). (B) Western blot analysis of total CD81 protein expression in WT Hepa1-6 and CD81KOH16 cells. GAPDH was used as loading control. (C-E) WT Hepa1-6 and CD81KOH16 cells were infected with PbGFP sporozoites and analyzed 24 h after invasion by flow cytometry (C) or microscopy (D, E) after staining with anti-UIS4 antibodies (red) and Hoechst 33342 nuclear stain (blue). The mean control values for each experiment were 0.27 and 0.93% PbGFP-infected cells (C), and 144, 145, 215 and 288 EEFs/well (D). ****p < 0.0001 (ratio paired t test). The images show PbGFP EEFs (green) surrounded by a UIS4-positive PV membrane (red) or intranuclear parasites in CD81KOH16 cells. Scale bar, 10 µm. Image collected and cropped by CiteAb from the following publication (https://pubmed.ncbi.nlm.nih.gov/32782257), licensed under a CC BY license.



Goat IgG (H+L) Cross-Adsorbed Secondary Antibody (A-11058) in ICC/IF
Identification of two hepatoblast subpopulations.a t-SNE plots showing the developmental stages (left) and clusters (right) of human (H) and mouse (M) endoderm-derived cells. b t-SNE plots showing the distinct clusters of ID3+ and ID3- hepatoblasts in the W5 human (H-W5) and E11.5 mouse (M-E11.5) fetal livers. c The proportion of ID3+ cells in human (H) and mouse (M) hepatoblasts /hepatocytes at different developmental time points. d Differentially expressed genes in W5 human (H-W5) and E11.5 mouse (M-E11.5) ID3+ and ID3- hepatoblasts. Each column represents a cell type and each row represents a gene. The TFs associated with each cell type are listed on the right. The color scheme is the same as b. e t-SNE plots showing the expression levels of marker genes. f Immunofluorescence showing the expression and distribution of ID3 and HNF4A in the W5 human (H-W5) and E11.5 mouse (M-E11.5) fetal livers. The arrowheads indicate ID3+ hepatoblasts. Scale bars, 60 μm. g The morphology of cultured hepatocytes (after 6-day culture from NCAM1+DLK+ hepatoblasts) and cholangiocyte tissue (after 10-day culture from NCAM1+DLK+ hepatoblasts). Scale bars, 20 μm. h Immunofluorescence showing the expression and distribution of HNF4A and SOX9 in cultured hepatocytes (upper) and cholangiocytes (lower), respectively. Scale bars, 15 μm. Image collected and cropped by CiteAb from the following publication (https://pubmed.ncbi.nlm.nih.gov/32690901), licensed under a CC BY license.

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1090 References

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